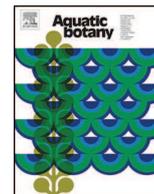




Contents lists available at ScienceDirect

## Aquatic Botany

journal homepage: [www.elsevier.com/locate/aquabot](http://www.elsevier.com/locate/aquabot)

# A comparison of photosynthetic and respiration rates in six aquatic carnivorous *Utricularia* species differing in morphology



Lubomír Adamec\*

Institute of Botany of the Academy of Sciences of the Czech Republic, Section of Plant Ecology, Dukelská 135, CZ-379 82 Třeboň, Czech Republic

## ARTICLE INFO

## Article history:

Received 9 December 2012  
 Received in revised form 3 June 2013  
 Accepted 11 June 2013  
 Available online 10 July 2013

## Keywords:

Lentibulariaceae  
 Linear and rhizomatous shoots  
 Oxygen-based net photosynthesis  
 Dark respiration  
 CO<sub>2</sub> uptake affinity  
 Shoot age  
 Metabolic costs  
 Growth traits

## ABSTRACT

The rootless carnivorous plant genus *Utricularia* includes about 50 submerged aquatic or amphibious species. They usually grow in standing humic waters rich in CO<sub>2</sub> and are known to be strict photosynthetic CO<sub>2</sub> users. In this study, oxygen-based net photosynthetic (P<sub>N</sub>) and dark respiration (RD) rates were measured in trap-free leaves of shoot segments from different aged sections of *Utricularia australis* and *U. vulgaris* and the photosynthetic CO<sub>2</sub> affinity was estimated in adult shoot segments. P<sub>N</sub> and RD were compared in adult trap-free leaves in three species with linear shoots (*U. australis*, *U. vulgaris*, *U. purpurea*) and four populations of three rhizomatous/rosette species (*U. dichotoma*, *U. resupinata*, *U. volubilis*) under standard conditions (25 °C, 400 μmol m<sup>-2</sup> s<sup>-1</sup> PAR, 0.25 mM CO<sub>2</sub>). In *U. australis* and *U. vulgaris* shoots, dry-weight based RD was highest in the youngest segments (174–184 mmol kg<sup>-1</sup> h<sup>-1</sup>) and gradually declined down to the progressively senescent segments (43–58 mmol kg<sup>-1</sup> h<sup>-1</sup>). In *U. vulgaris*, P<sub>N</sub> increased from the shoot apex up to the 25th leaf nodes, while it was highest in the 3rd leaf nodes and decreased significantly down to the shoot bases in *U. australis*. Chlorophyll-based P<sub>N</sub> rates along the shoots in both species were almost constant. Dry-weight based P<sub>Nmax</sub> in *U. australis* was significantly greater than that in *U. vulgaris* (1634 ± 69 vs. 1077 ± 56 mmol kg<sup>-1</sup> h<sup>-1</sup>), while the CO<sub>2</sub> affinity (K<sub>m</sub>) was opposite – 48 ± 10 vs. 21 ± 2 μM. Although the mean photosynthetic parameters for individual species usually differed considerably from each other between both subgroups and supported the view that the linear-shoot species have higher P<sub>N</sub> rates than the other subgroup (732–1592 vs. 155–687 mmol kg<sup>-1</sup> DW h<sup>-1</sup>), statistically significant difference between both subgroups was only found for chlorophyll *a*-based P<sub>N</sub>. The differences in photosynthetic characteristics found between both morphological subgroups of *Utricularia* presumably reflect the differences in growth rates: the linear-shoot species with high P<sub>N</sub> rates are known to grow very rapidly while those from the other subgroup with lower P<sub>N</sub> grow slowly. The high P<sub>N</sub> rates of the linear-shoot *Utricularia* species approach the upper limit of P<sub>N</sub> reported for other aquatic non-carnivorous plants and are a prerequisite for their very rapid growth. Therefore, to attain such high P<sub>N</sub> rates and rapid growth, these species demand high [CO<sub>2</sub>] > 0.15 mM in their habitats.

© 2013 Elsevier B.V. All rights reserved.

## 1. Introduction

The rootless carnivorous plant genus *Utricularia* L. (bladderwort, Lentibulariaceae) includes about 220 species which are subdivided into 35 generic sections (Taylor, 1989; Jobson et al., 2003). The majority of the species are wetland, terrestrial ones but about 50 are submerged aquatic or amphibious species belonging mainly to the *Utricularia*, *Pleiochasia*, *Lectricula*, *Avesicaria*, *Avesicarioides* and *Vesiculina* sections. The typical aquatic or amphibious species from the *Utricularia* and *Vesiculina* sections have a linear, modular and fairly

regular shoot structure, consisting of nodes with dissected leaves reminiscent of whorls and thin cylindrical internodes (Friday, 1989; Taylor, 1989; Sattler and Rutishauser, 1990; Rutishauser, 1993). The aquatic or amphibious species from the other sections have rhizomatous shoot structures with flat, lanceolate or narrow cylindrical leaves growing in nodes (rarely as rosettes) from short filiform stems (Taylor, 1989). In this way, they are reminiscent of the terrestrial *Utricularia* species. In this paper, I shall use the term “leaf” for the dissected or flat organ with the prevailing photosynthetic function, growing in leaf nodes (or untrue whorls) in a plane perpendicular to the “stem” (stolon).

Aquatic *Utricularia* species usually grow in oligo-mesotrophic, humic (dystrophic) waters with a high concentration of free CO<sub>2</sub> commonly exceeding 0.1–0.2 mM and all aquatic species tested so far use only CO<sub>2</sub> for photosynthesis (Moeller, 1978; Adamec, 1997a, 2007, 2008a,b, 2009, 2011a, 2012; Adamec and Kovářová, 2006; Pagano and Titus, 2007; Adamec and Pásek, 2009). Generally, in 12

Abbreviations: DR, dark respiration rate; P<sub>N</sub>, net photosynthetic rate; P<sub>Nmax</sub>, maximum net photosynthetic rate; chl. *a*, chlorophyll *a*; DW, dry weight; FW, fresh weight; K<sub>m</sub>, half-saturation constant.

\* Tel.: +420 384 721 156; fax: +420 384 721 136.

E-mail addresses: [adamec@butbn.cas.cz](mailto:adamec@butbn.cas.cz), [lubo.adamec@seznam.cz](mailto:lubo.adamec@seznam.cz)

aquatic *Utricularia* species with linear shoots (sections *Utricularia* and *Vesiculina*) growing in the field or semi-natural cultures, the CO<sub>2</sub> compensation point of photosynthesis fell within the range 1.2–13.6 μM (mean 3–5 μM) and was the same (mean 5.2 μM, range 2.5–8.8 μM) for these species growing *in vitro* (Moeller, 1978; Adamec, 1997a, 2008b, 2009, 2011a; Adamec and Kovářová, 2006; Pagano and Titus, 2007; Adamec and Pásek, 2009). Similar values of 1.5–10 μM are reported in many aquatic non-carnivorous plants (Maberly and Spence, 1983). All results show clearly that the CO<sub>2</sub> compensation point in aquatic *Utricularia* species is rather variable and adaptable.

The very rapid growth of most of aquatic *Utricularia* species with linear shoots, even in oligotrophic habitats, is associated with both a high apical shoot growth rate of 2–4 leaf nodes per day and frequent vegetative propagation by branching (Friday, 1989; Adamec, 2008b, 2009, 2010, 2011a,b, 2012; Ellison and Adamec, 2011). The rapid growth of aquatic *Utricularia* species requires several ecophysiological adaptations: high photosynthetic rate of shoots, prey capture, efficient nutrient re-utilization from senescent shoots and high nutrient uptake affinity from the ambient water (Richards, 2001; Adamec, 2008a,b, 2009, 2011a, 2012). Aquatic *Utricularia* species in their habitats may also face shortage of light (only 2–20% of incident PAR irradiance) and sometimes also free CO<sub>2</sub> (<0.02–0.05 mM; Adamec, 1997a, 2008a, 2009, 2012). Therefore, their net photosynthetic rate (P<sub>N</sub>) in standing waters, like that of other submerged plants generally, is limited by unfavorable physical and chemical factors. They include a low diffusion rate of CO<sub>2</sub>, variable [CO<sub>2</sub>] strongly dependent on pH and total alkalinity (TA) and a shortage of light (e.g., Maberly and Madsen, 2002). Even though available data are limited, the maximum P<sub>N</sub> (P<sub>Nmax</sub>) of aquatic *Utricularia* with linear shoots (eight species, 40–148 mmol O<sub>2</sub> kg<sup>-1</sup> (fresh weight) h<sup>-1</sup> or 650–1860 mmol O<sub>2</sub> kg<sup>-1</sup> (dry weight) h<sup>-1</sup>) is usually comparable with the highest values of P<sub>Nmax</sub> found in aquatic non-carnivorous species (ca. 300–1100 mmol O<sub>2</sub> kg<sup>-1</sup> DW h<sup>-1</sup>) or higher (cf. Moeller, 1978; Pokorný and Ondok, 1991; Madsen et al., 1993; Maberly and Madsen, 2002; Kahara and Vermaat, 2003; Adamec, 2006, 2008b, 2011a,c; Ellison and Adamec, 2011). Nevertheless, the P<sub>N</sub> values in aquatic *Utricularia* shoots/leaves measured under natural habitat conditions or in natural water may be considerably lower (*U. vulgaris*: 120–380 mmol kg<sup>-1</sup> DW h<sup>-1</sup>, Draxler, 1973; *U. macrorhiza*: 70–500 mmol kg<sup>-1</sup> DW h<sup>-1</sup>, Knight, 1992).

Traps of aquatic *Utricularia* species are physiologically very active organs having high aerobic dark respiration rates (RD) and large photosynthetic and energetic (maintenance) costs (Knight, 1992; Adamec, 2006, 2011a). In six species, trap RD (90–153 mmol kg<sup>-1</sup> DW h<sup>-1</sup>) was 1.7–3.0 times greater than that in leaves on carnivorous or photosynthetic shoots, while trap P<sub>Nmax</sub> (112–264 mmol kg<sup>-1</sup> DW h<sup>-1</sup>) was 7–10 times lower than that in photosynthetic leaves (Adamec, 2006). This very high RD:P<sub>Nmax</sub> ratio in traps (50–140%) is not found in leaves (only 3.6–8.2%) and means that there are high maintenance costs but very low photosynthetic efficiency in traps. In spite of a steep physiological polarity along the shoots of aquatic *Utricularia* in tissue N and P content (Adamec, 2008a; and carbohydrate content in ecologically similar *Aldrovanda vesiculosa*; Adamec, 2000), the measurements of P<sub>N</sub> and RD in *U. macrorhiza* shoot segments of different age from the apex to the senescent, trap-free segments showed a virtually constant P<sub>N</sub> (DW- or chlorophyll-based) along the whole shoot (Knight, 1992).

In conclusion, very high P<sub>N</sub> of shoots is typical for aquatic *Utricularia* species with linear shoots and rapid growth. It is also a prerequisite for this rapid growth as the rapid decay of senescent shoot segments causes a great and permanent loss of structural and non-structural carbohydrates together with the high maintenance costs of traps (Adamec, 2011a). However, photosynthetic and growth traits of aquatic (amphibious) rhizomatous *Utricularia*

species from the *Pleiochasia* and *Lecticula* sections have never been studied. The aim of this study has been to compare P<sub>N</sub> values (DW-, FW-, and chlorophyll *a*-based) in three species of aquatic *Utricularia* with homogeneous, linear shoots (*U. vulgaris*, *U. australis*, *U. purpurea*) with those in three aquatic/amphibious species with rhizomatous or rosette-shaped shoots (*U. dichotoma*, *U. resupinata*, *U. volubilis*). As opposed to rapid growth of the species from the first group (Adamec, 2011a), those from the second group grow rather slowly in cultures (Adamec, unpubl.). It is thus possible to test whether photosynthetic traits relate to growth traits within aquatic *Utricularia* species. For two species from the first group (*U. vulgaris*, *U. australis*), P<sub>N</sub> and RD were measured as standard in trap-free leaves from the apex down to the senescent shoot basis, to estimate a photosynthetic shoot polarity and CO<sub>2</sub> uptake affinity (K<sub>m</sub>) was estimated in mature shoot segments.

## 2. Materials and methods

### 2.1. Plant material

Measurements were performed in three aquatic *Utricularia* species with homogeneous, linear shoots (*U. vulgaris* L., *U. australis* R.Br. /both collected from the Czech Republic/ – section *Utricularia*, and *U. purpurea* Walt. /from Florida, USA/ – section *Vesiculina*) and in three aquatic/amphibious species with rhizomatous or rosette shoots (two distinct populations of *U. dichotoma* Labill. /collected from Newcastle, N.S.W., Australia, with robust leaves, and from Katoomba, N.S.W., Australia, with normal-sized leaves/, *U. volubilis* R.Br./rosette, from W.A., Australia/ – all of them section *Pleiochasia*, and *U. resupinata* Greene ex Bigelow /from Nicaragua/ – section *Lecticula*). *U. vulgaris* and *U. australis* were grown outdoors in two plastic containers (area 0.6–3 m<sup>2</sup>, 200–2000 L) in which the water chemistry, light and temperature conditions were similar to natural conditions. *U. purpurea* and *U. volubilis* were grown in a naturally lit greenhouse in a 300 L plastic container. The plants in these cultures were grown in tap water and a litter of robust *Carex* species was used as a substrate. The pH of the cultivation media was 6.9–7.5, dissolved oxygen concentration ranged from 0.15 to 0.30 mM, total alkalinity from 0.6 to 0.9 mequiv. L<sup>-1</sup> and the free CO<sub>2</sub> concentration was 0.08–0.20 mM (for all details, see Adamec, 1997b, 2008b). The water in these cultures was considered oligotrophic (in μg L<sup>-1</sup>: NO<sub>3</sub><sup>-</sup>-N, 0–12; NH<sub>4</sub><sup>+</sup>-N, 8–16; PO<sub>4</sub>-P, 19–27) and humic. Both populations of *U. dichotoma* and *U. resupinata* were grown in the greenhouse as submerged plants in 3 L aquaria floating in the 300 L container and a mixture of peat with sand was used as the substrate. The pH of the cultivation media was 6.1–6.2. All outdoor and greenhouse cultures were partly shaded. Small zooplankton species were added weekly to most cultures to promote plant growth. Adult plants of all species were used for measurements from 18 July to 8 August.

### 2.2. Gas exchange measurements

For *U. vulgaris* and *U. australis*, RD and P<sub>N</sub> were measured on 2–4 freshly collected leaves (from 1 to 2 plants) of five different ages (shoot segments) which were thoroughly stripped of mature traps. *U. vulgaris* plants were 55–70 cm long with 72–98 adult leaf nodes and *U. australis* were 60–80 cm long with 76–88 leaf nodes. They were very young, subapical leaves 5–8 mm long positioned just below the shoot apex bearing immature traps 0.5–1 mm large and adult leaves (without traps) from the 3rd, 10th, 25th and 50–60th adult leaf nodes. The leaves from the 3rd, 10th and 25th leaf nodes bore mature functional traps, while the leaves from the 50th to 60th leaf nodes were partly senescent and usually trap-free. Except for the youngest subapical leaves, all traps on all older leaves were

thoroughly removed using two pairs of forceps as the trap  $P_N$  is very low and the trap proportion to the shoot biomass may be variable along shoots (Friday, 1989; Adamec, 2006, 2008a). In these two species, the dependence of  $P_N$  on  $[CO_2]$  was also measured in trap-free leaves from the 10th leaf nodes. The measured leaves were exposed successively to new media with 0.02, 0.05, 0.10, 0.25, 0.50 and 1.00 mM free  $CO_2$  prepared by bubbling with  $CO_2$  according to pH (Helder, 1988). To compare photosynthetic traits in various species, RD and  $P_N$  were measured in 3–4 leaves (with traps removed) from the 2nd adult leaf nodes of *U. purpurea*, 2–7 mature leaves of *U. dichotoma* (of both populations), 14–18 mature leaves of *U. resupinata* (3–6 cm long), and 8–12 mature leaves (6–15 cm long) of *U. volubilis*. The leaves of *U. resupinata* were cut into 1.5–2 cm long segments. The large air spaces in these were filled with the experimental solution using negative pressure in a syringe before the measurements. After this treatment, only a minor part of fine intercellulars was not filled by water. *U. volubilis* leaves were cut to 2–3 cm segments. FW of measured leaves of all species was 5–46 mg.

Generally for all measurements, RD and  $P_N$  were measured in a solution of 1 mM  $NaHCO_3$  with 0.1 mM KCl (ca. 90%  $O_2$  saturation) in a 4.2- or 5.3-mL stirred chamber kept at  $25.0 \pm 0.1^\circ C$ . A Clark-type oxygen sensor and a pen recorder (for details see Adamec, 1997b, 2006) was used. Free  $[CO_2]$  of 0.25 mM was used as standard where possible. This  $[CO_2]$  was chosen as it has been used commonly in many similar studies (cf. Adamec, 1997b, 2006, 2008b, 2011c), it often occurs in natural habitats and presumably approaches that for achieving the  $P_{Nmax}$ . After RD had been measured in darkness for 12–15 min, a light was switched on (halogen reflector,  $400 \mu mol m^{-2} s^{-1}$  PAR) and  $P_N$  was measured for 12–15 min. FW was then estimated for all measured leaves and chlorophyll *a* content was determined (Pechar, 1987) after the measurements of shoot polarity in *U. vulgaris* and *U. australis* and the comparative interspecific measurements. DW ( $80^\circ C$ ) was estimated in parallel foliar samples. All measurements were repeated six times under the same conditions using material from different plants. RD and  $P_N$  are expressed in  $mmol kg^{-1} DW h^{-1}$  as standard.

### 2.3. Statistical treatment

The plot of  $P_N$  against  $[CO_2]$  for single measurements was fitted by a nonlinear regression. The Michaelis-Menten model, modified to include DR, was used to fit the data to obtain  $K_m$  and  $P_{Nmax}$  parameters. The statistically significant differences in these parameters between *U. vulgaris* and *U. australis* were tested using a two-tailed *t*-test. In these species, 1-way ANOVA was used to find significant differences in RD and photosynthetic parameters between the neighboring shoot segments to prove the physiological polarity along the shoots. Significant differences in RD and photosynthetic parameters within the three species with linear shoots, as well as within the three species with rhizomatous shoots, were found using 1-way ANOVA. Two-way ANOVA was used to find significant differences between these two morphological *Utricularia* groups. Species, nested within a morphological group, were chosen as a factor with random effect. Significant correlations between RD and  $P_N$  were sought using linear regressions of the data. All statistical analyses were performed using STATISTICA v. 10 (StatSoft, USA). Throughout the paper, means  $\pm 1$  SE are shown.

## 3. Results

In *U. australis* and *U. vulgaris* trap-free leaves from the 10th adult leaf nodes, the plot of  $P_N$  against  $[CO_2]$ , modified to include DR, corresponded to the Michaelis-Menten model (Fig. 1; coefficient of determination for pooled data 0.92 and 0.85, respectively).

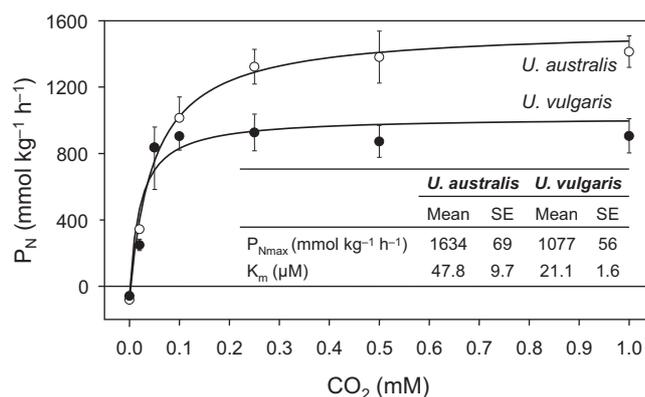


Fig. 1.  $P_N$  rates (DW-based) in trap-free leaves from the 10th adult leaf nodes of *U. australis* and *U. vulgaris* as dependent on  $CO_2$  concentration. Pooled data were fitted by the Michaelis-Menten model including RD. The negative values at zero  $[CO_2]$  denote RD. Means  $\pm$  SE intervals are shown;  $n = 6$ .  $P_{Nmax}$ , calculated maximum net photosynthetic rate;  $K_m$ , half-saturation constant.

$P_{Nmax}$  in *U. australis* was significantly greater ( $p < 0.002$ ,  $t = 5.24$ ) than that in *U. vulgaris* ( $1634 \pm 69$  vs.  $1077 \pm 56$   $mmol kg^{-1} DW h^{-1}$ ), while the  $CO_2$  affinity ( $K_m$ ) was opposite:  $48 \pm 10$  vs.  $21 \pm 2$   $\mu M$  ( $p < 0.05$ ,  $t = 2.72$ ). In both species,  $P_{Nmax}$  was attained at only about 0.25 mM  $CO_2$ . As it follows from the comparison of photosynthetic traits in trap-free leaves along the shoots, chlorophyll *a* content in all tested shoot segments in *U. australis* exceeded that in *U. vulgaris* by 1.8–2.7 times (Table 1). RD values were comparable in both species. In both species, RD was the highest in the youngest, sub-apical segment and gradually declined down to the progressively senescent segments. Yet RD in the old, trap-free shoot segments (50–60th leaf nodes) was the same as that in the 25th leaf nodes with still fully functional leaves and traps. The course of DW-based  $P_N$  along the shoots was different in both species: in *U. vulgaris*,  $P_N$  increased from the shoot apex up to the 25th leaf node, while it was the highest in the 3rd leaf nodes and decreased significantly down to the shoot bases in *U. australis*. The same pattern held also for FW-based  $P_N$  (data not shown). The course of chlorophyll-based  $P_N$  along the shoots was similar in both species, more or less constant along the adult shoot segments, although there was a highly significant decline of chlorophyll *a* content in the senescent segments. The values were similar in both species. The RD: $P_N$  ratio as a measure of photosynthetic efficiency was significantly ( $p < 0.01$ ,  $F_{4,25} > 15.2$ ) the highest in the subapical leaves and declined to the 25th leaf nodes in both species but the photosynthetic efficiency was significantly better in *U. australis* (Table 1).

From the comparison of photosynthetic traits between three aquatic *Utricularia* species with linear shoots, *U. australis* had significantly ( $p < 0.05$ ,  $F_{2,11} > 45.9$ ) the highest chl. *a* content and the DW- and FW-based  $P_N$  rates, while the chl. *a*-based  $P_N$  rates were comparable (Table 2). This species also had the highest photosynthetic efficiency. Generally, within this subgroup of three species,  $P_N$  ranged between  $732$ – $1592$   $mmol kg^{-1} DW h^{-1}$ ,  $61$ – $145$   $mmol kg^{-1} FW h^{-1}$ , or  $194$ – $235$   $mmol g^{-1} chl. a h^{-1}$ . Within the subgroup of three aquatic *Utricularia* species with rhizomatous or rosette shoots, chl. *a* content in leaves ranged between  $4.8$ – $13.0$   $g kg^{-1} DW$  and was the highest in robust *U. dichotoma* from Newcastle, N.S.W. (Table 2). Within this subgroup,  $P_N$  ranged between  $155$ – $687$   $mmol kg^{-1} DW h^{-1}$ ,  $5.7$ – $63$   $mmol kg^{-1} FW h^{-1}$ , or  $28$ – $79$   $mmol g^{-1} chl. a h^{-1}$ . The species with the lowest  $P_N$  rates and efficiency were *U. resupinata* and the robust *U. dichotoma* (Newcastle), while the rosette species *U. volubilis* and rhizomatous *U. dichotoma* from Katoomba, N.S.W., had the highest  $P_N$  rates and efficiency. Due to distinctly different DW proportion between both subgroups, the comparison of FW-based  $P_N$  or RD rates does not

**Table 1**  
Oxygen-based RD and P<sub>N</sub> rates of trap-free leaves of *Utricularia vulgaris* and *U. australis* of different age.

Leaf age (adult nodes)	<i>Utricularia vulgaris</i>										<i>Utricularia australis</i>									
	DW (% FW)	Chl. <i>a</i> (g kg <sup>-1</sup> DW)	RD <sub>DW</sub> (mmol kg <sup>-1</sup> h <sup>-1</sup> )	P <sub>NDW</sub> (mmol kg <sup>-1</sup> h <sup>-1</sup> )	P <sub>Nchl</sub>	RD/P <sub>N</sub>	DW (% FW)	Chl. <i>a</i> (g kg <sup>-1</sup> DW)	RD <sub>DW</sub> (mmol kg <sup>-1</sup> h <sup>-1</sup> )	P <sub>NDW</sub> (mmol kg <sup>-1</sup> h <sup>-1</sup> )	P <sub>Nchl</sub>	RD/P <sub>N</sub>								
Apex	8.75	2.23 ± 0.12	184** ± 8.3	362** ± 9.4	155 ± 5.6	0.521** ± 0.044	8.37	6.10* ± 0.37	174* ± 27	1158** ± 56	191** ± 5.6	0.155** ± 0.026								
3rd	10.3	3.33 ± 0.06	86.9 ± 8.0	739 ± 36	222 ± 11	0.119 ± 0.011	8.53	7.46 ± 0.25	103 ± 13	1855 ± 62	248 ± 4.7	0.056 ± 0.008								
10th	10.7	3.39 ± 0.16	65.4 ± 9.9	732 ± 78	216 ± 23	0.109 ± 0.039	9.09	6.78 ± 0.07	59.9 ± 5.9	1592* ± 67	235 ± 10	0.038 ± 0.003								
25th	10.8	3.88** ± 0.32	44.2 ± 5.7	845* ± 81	218 ± 18	0.052 ± 0.010	8.38	7.09** ± 0.27	53.5 ± 3.5	1451** ± 63	205 ± 9.8	0.037 ± 0.004								
50–60th	10.9	2.39 ± 0.41	42.5 ± 4.0	538 ± 76	227 ± 23	0.084 ± 0.009	7.24	5.38 ± 0.41	58.1 ± 4.6	1119 ± 69	209 ± 14	0.053 ± 0.005								

RD and P<sub>N</sub> are expressed per unit DW, P<sub>N</sub> also per unit chlorophyll *a* in mmol g<sup>-1</sup> chl. *a* h<sup>-1</sup>. Percent of DW in FW and chlorophyll *a* content per DW are also shown. Means ± SE intervals are shown; *n* = 6. The asterisks denote statistically significant difference between the given and the next shoot segment (\**p* < 0.05; \*\**p* < 0.01; 1-way ANOVA).

reveal much. On the DW basis, RD in the linear-shoot species ranged between 55–65 mmol kg<sup>-1</sup> h<sup>-1</sup> and was the same as that in the latter subgroup (35–84 mmol kg<sup>-1</sup> h<sup>-1</sup>). The mean values of photosynthetic parameters for individual species usually differed considerably between both subgroups from each other and this supported the view that the linear-shoot species have higher P<sub>N</sub> rates than the other subgroup. Statistically significant differences between both subgroups were, however, only found for chl. *a*-based P<sub>N</sub> (*p* < 0.0006, *F*<sub>1,35</sub> = 59.6). The DW- and FW-based P<sub>N</sub> rates were only weakly different from each other (*p* < 0.057–0.063; Table 2). The linear regression of FW-based P<sub>N</sub> and RD revealed a statistically significant correlation only for all species (P<sub>N</sub> = 26.1 + 8.28 RD; *n* = 36, *r*<sup>2</sup> = 0.132, *p* = 0.029) and the rhizomatous/rosette species (P<sub>N</sub> = -3.70 + 10.0 RD; *n* = 18, *r*<sup>2</sup> = 0.309, *p* = 0.016), but not for the linear-shoot species (data not shown).

#### 4. Discussion

In this study, photosynthetic traits were compared in three aquatic *Utricularia* species with linear shoots which have often been used for ecophysiological and photosynthetic research (e.g., Moeller, 1978, 1980; Friday, 1989; Richards, 2001; Adamec, 2006, 2008a,b, 2009, 2011a,c; Adamec and Kovářová, 2006; Pagano and Titus, 2007; Adamec and Pásek, 2009), while the other species with rhizomatous or rosette shoots – *U. dichotoma*, *U. resupinata* and *U. volubilis* – have never been studied in this respect and their ecophysiology is quite unknown. The linear-shoot species *U. australis* and *U. vulgaris* from the section *Utricularia* are known for rapid apical growth with a high relative growth rate (0.9–4.2 leaf nodes d<sup>-1</sup>; Friday, 1989; Adamec and Kovářová, 2006; Adamec, 2008b, 2009, 2011a), while *U. purpurea* (section *Vesiculina*) grows much slower (0.25 leaf nodes d<sup>-1</sup>; Richards, 2001; see also Moeller, 1980). Although a close correlation between the relative growth rate and the DW- or leaf area-based P<sub>Nmax</sub> generally holds for terrestrial plants (Shipley, 2006), it has not been found within the subgroup of three aquatic *Utricularia* species (Table 2). P<sub>N</sub> values of slowly growing *U. purpurea* were similar to those of rapidly growing *U. vulgaris* and the same held for the RD:P<sub>N</sub> ratio. The former species may invest more photosynthates either to trap exudation (cf. Sirová et al., 2009, 2010) or to the thick gelous slime covering its shoots.

In both *U. vulgaris* and *U. australis*, P<sub>N</sub> measurements in trap-free leaves in shoot segments of different age revealed relatively high P<sub>N</sub> values in the oldest (50–60th) segments. This corresponded to the significantly decreased chl. *a* content (Table 1); the FW-based P<sub>N</sub> rates reached about 51–64% of the maximum P<sub>N</sub> in younger segments, while the chl. *a*-based P<sub>N</sub> rates were nearly constant. A similar pattern of chl. *a* content and chl. *a*-based P<sub>N</sub> was also found in shoot segments of different age of *U. macrorhiza* (Knight, 1992; very relative to *U. vulgaris*). It is thus possible to conclude that aquatic *Utricularia* species from the generic *Utricularia* section with linear, homogeneous shoots and rapid apical growth dispose of high P<sub>N</sub> rates even in very old, trap-free shoot segments. This contributes to covering both great photosynthetic and energetic (maintenance) costs associated with production and activity of traps (Knight, 1992; Adamec, 2006, 2011a; Sirová et al., 2010, 2011; Borovec et al., 2012) and requirements associated with rapid growth (allocation of carbohydrates to new biomass, great loss of biomass in basal decaying shoot segments; see Adamec, 1997b for similar *A. vesiculosa*). As recently shown by Borovec et al. (2012) exudation of organic matter into traps to support commensal communities depends on the photosynthetic conditions of *Utricularia* plants.

Some ecological differences between *U. vulgaris* and *U. australis* clearly follow from the present results. In all shoot segments from

**Table 2**  
Oxygen-based RD and P<sub>N</sub> rates of adult trap-free leaves of 6 aquatic *Utricularia* species.

Species	DW (% DW)	Chl. <i>a</i> (g kg <sup>-1</sup> DW)	RD <sub>DW</sub> (mmol kg <sup>-1</sup> h <sup>-1</sup> )	P <sub>N</sub> DW	P <sub>N</sub> FW	P <sub>N</sub> chl (mmol g <sup>-1</sup> chl. h <sup>-1</sup> )	RD/P <sub>N</sub>
<i>U. australis</i>	9.09	6.78 ± 0.07a	59.9 ± 5.9a	1592 ± 67a	145 ± 6.1a	235 ± 10a	0.038 ± 0.003a
<i>U. vulgaris</i>	10.7	3.39 ± 0.16b	65.4 ± 9.9a	732 ± 78b	78.4 ± 8.4b	216 ± 23a	0.109 ± 0.040a
<i>U. purpurea</i>	7.36	4.30 ± 0.29c	55.1 ± 7.6a	835 ± 60c	61.4 ± 4.4c	194 ± 7.0a	0.070 ± 0.012a
<i>U. dichotoma</i> <sup>a</sup>	5.73	13.0 ± 0.28a	82.7 ± 6.1a	359 ± 31a	20.6 ± 1.8a	27.8 ± 2.7a	0.241 ± 0.048ab
<i>U. dichotoma</i> <sup>b</sup>	5.31	10.0 ± 0.69b	83.7 ± 6.2a	687 ± 68b	36.5 ± 3.6b	70.4 ± 9.1b	0.130 ± 0.011ac
<i>U. resupinata</i>	3.65	4.37 ± 0.40c	44.1 ± 4.7b	155 ± 14c	5.65 ± 0.51c	36.8 ± 4.6a	0.283 ± 0.058b
<i>U. volubilis</i>	12.4	6.45 ± 0.21d	35.1 ± 2.0b	509 ± 46a	62.9 ± 5.7d	78.9 ± 6.5b	0.071 ± 0.006c
<i>F</i> <sub>1,35</sub>	–	2.13	2.47	5.65	6.08	59.6	3.41
<i>p</i>	–	0.20	0.18	0.063	0.057	0.0006	0.12

<sup>a</sup> Robust *U. dichotoma* from Newcastle, N.S.W.

<sup>b</sup> *U. dichotoma* population from Katoomba near Sydney, N.S.W.

RD and P<sub>N</sub> are expressed per unit DW, P<sub>N</sub> also per unit FW and chlorophyll *a* in mmol g<sup>-1</sup> chl. a h<sup>-1</sup>. *U. resupinata* leaves were infilled by water. Percent of DW in FW and chlorophyll *a* content per unit DW are also shown. Means ± SE intervals are shown; *n* = 6. The species with linear shoots are separated by a dotted line from the species with stoloniferous or rosette shoots. At the bottom, significant difference between both subgroups is shown (nested-design of ANOVA). The different letters within each plant subgroup denote statistically significant differences between species (*p* < 0.05; 1-way ANOVA).

the same position and of similar age, FW- and DW-based P<sub>N</sub> values were significantly higher in *U. australis* (Table 1) and also the P<sub>N</sub>max in adult shoot segments was 41% higher than that in *U. vulgaris* (Fig. 1). In both species, the P<sub>N</sub> rate at the standard [CO<sub>2</sub>] of 0.25 mM approached the P<sub>N</sub>max. On the other hand, CO<sub>2</sub> affinity in *U. australis* was about twice as low as in *U. vulgaris*. Higher P<sub>N</sub>max and the photosynthetic efficiency in *U. australis* confirm its higher apical shoot growth rate when compared with *U. vulgaris* (cf. Friday, 1989; Adamec, 2008b, 2009, 2011a; Adamec and Kovářová, 2006). However, the much lower CO<sub>2</sub> affinity in *U. australis* suggests that this species is primarily adapted to much higher CO<sub>2</sub> concentrations (and to much lower pH values) in the water (commonly peatbogs) than *U. vulgaris*, which prefers hard and slightly alkaline waters with lower [CO<sub>2</sub>] (Adamec, 1997a, 2008a, 2009).

Generally, the three *Utricularia* species with rhizomatous or rosette shoots used in this study resemble typical terrestrial *Utricularia* species as they have similarly elongated, flat or tubular leaves 0.4–4 mm wide with a relatively low surface:volume ratio, as opposed to the linear-shoot species with fine filamentous leaves (Taylor, 1989). All three species grow slowly in cultures (Adamec, unpubl.). Moreover, *U. dichotoma* and *U. resupinata* grow in very shallow waters and may be considered true amphibious species (Taylor, 1989). Both populations of *U. dichotoma* had a significantly higher chl. *a* content in leaves than the other two species from the subgroup, while that in *U. resupinata* and *U. volubilis* was comparable with those in the linear-shoot species (Table 2). Overall, the DW- and chl. *a*-based P<sub>N</sub> values and also the photosynthetic efficiency were somewhat higher in the linear-shoot species than in those from the other subgroup but, due to the small number of species studied and the relatively high variance of the results, the difference was significant only in chl. *a*-based P<sub>N</sub>. Similarly, the comparison of photosynthetic traits between two functional subgroups of aquatic *Utricularia* species shows that some photosynthetic parameters clearly overlap between both subgroups thus forming a rather gradual transition of these parameters between the subgroups than a qualitative, sharp distinction. Within all (including the rhizomatous/rosette) species used, the discovered correlation between higher P<sub>N</sub> rates and higher RD rates in trap-free leaves suggests a generally higher metabolic activity in leaves in some species. The linear-shoot species used in this study are known to have a relatively high proportion of traps to the total plant biomass as an investment in carnivory (4–62%; Friday, 1989; Adamec, 2008a, 2009). In the rhizomatous species used, an unknown but significant biomass proportion contains subterranean pale rhizomes or rhizoids as non-photosynthetic organs (Taylor, 1989).

In this study, a relatively high CO<sub>2</sub> affinity was found in *U. vulgaris* and *U. australis* trap-free leaves (*K*<sub>m</sub> 21 and 48 μM; Fig. 1).

However, in carnivorous *A. vesiculosa*, which commonly accompanies both species, *K*<sub>m</sub> was 165 μM (Adamec, 1997b) and in *U. purpurea* shoots collected from natural sites in the USA, CO<sub>2</sub> affinity was even markedly lower (*K*<sub>m</sub> about 1.0–1.7 mM; Moeller, 1978). The CO<sub>2</sub> compensation points of photosynthesis found for 12 linear-shoot aquatic *Utricularia* species at many field sites or in nearly natural cultures or experiments are, however, consistently within a relatively narrow range of 1.2–13.6 μM (Moeller, 1978; Adamec, 1997a, 2008b, 2009, 2011a; Adamec and Kovářová, 2006; Pagano and Titus, 2007; Adamec and Pásek, 2009) and resemble those commonly reported in aquatic plants (Maberly and Spence, 1983; Sand-Jensen and Frost-Christensen, 1999).

The P<sub>N</sub>max approaching values measured under standard laboratory conditions for three linear-shoot *Utricularia* species (Table 2) are the same or much higher than those reported by various authors for leaves or shoots for 8 aquatic *Utricularia* species with linear shoots (range ca. 8–148 mmol kg<sup>-1</sup> FW h<sup>-1</sup> or 80–1860 mmol kg<sup>-1</sup> DW h<sup>-1</sup>; Draxler, 1973; Moeller, 1978; Knight, 1992; Adamec, 2006, 2011a). Besides, high P<sub>N</sub> values of 150–1050 mmol kg<sup>-1</sup> DW h<sup>-1</sup> were also found in old or new segments of sprouting turions of 4 aquatic *Utricularia* species (Adamec, 2011c). Chlorophyll *a* content found in both subgroups of aquatic *Utricularia* (3.4–13 g kg<sup>-1</sup> DW; Table 2) is also the same or comparable to that reported for *U. vulgaris* and *U. macrorhiza* shoots (2–12 g kg<sup>-1</sup> DW; Maier, 1973; Knight, 1992) and/or for many aquatic plants (3–18 g kg<sup>-1</sup> DW; Madsen et al., 1993; Adamec, 2000; Kahara and Vermaat, 2003). Overall, if the P<sub>N</sub>max of the majority of aquatic plant species usually reaches only about 300–1100 mmol kg<sup>-1</sup> DW h<sup>-1</sup> or 30–110 mmol kg<sup>-1</sup> FW h<sup>-1</sup> (for review see Pokorný and Ondok, 1991; Madsen et al., 1993; Maberly and Madsen, 2002; Kahara and Vermaat, 2003), it indicates that P<sub>N</sub> values (both on DW, FW and chlorophyll basis) found in many aquatic *Utricularia* species with linear shoots approach an upper limit of P<sub>N</sub>max reported for all other aquatic plants (cf. also Maberly, 1985).

In conclusion, aquatic *Utricularia* species usually grow in waters with high CO<sub>2</sub> concentrations (>0.15 mM; Adamec, 2007, 2008a, 2011a) and under sufficient light and temperature habitat conditions, their P<sub>N</sub> rates can be very high and approach the P<sub>N</sub>max. The crucial importance of [CO<sub>2</sub>] for attaining high growth rate was proven for 4 aquatic *Utricularia* species in which an increase in [CO<sub>2</sub>] in growth experiments led to a marked and significant increase in their relative growth rate (McDermott and Darnowski, 2002; Pagano and Titus, 2004, 2007); the same effect commonly also occurs in other aquatic non-carnivorous plants (e.g., Sand-Jensen and Frost-Christensen, 1999; Pagano and Titus, 2004, 2007). The same ecological strategy – growth dependence on CO<sub>2</sub>-rich waters

– occurs also in the rooted floating macrophyte *Stratiotes aloides* (Nielsen and Borum, 2008), which often accompanies *Utricularia* stands. Moreover, relatively high  $P_N$  is also kept in senescent shoot segments of *Utricularia*. Very high  $P_N$  of (photosynthetic) shoots of rapidly growing *Utricularia* species with linear shoots is therefore a prerequisite for this rapid growth. On the other hand, aquatic or amphibious *Utricularia* species with rhizomatous/rosette shoots do not require high  $P_N$  rates for their slow growth. As a result of their growing in very shallow waters as amphibious species, they can also take up atmospheric  $CO_2$  by their aerial leaves when they emerge.

### Acknowledgements

Sincere thanks are due to Dr. Brian G. McMillan for correction of the language and to Dr. Tomáš Hájek for technical help. Thanks are also due to Dr. Richard W. Jobson for providing two populations of *Utricularia dichotoma* for research. This study was partly supported by the Czech Research Project CSF P504/11/0783 and the Long-term research development project no. RVO 67985939.

### References

- Adamec, L., 1997a. Mineral nutrition of carnivorous plants: a review. *Bot. Rev.* 63, 273–299.
- Adamec, L., 1997b. Photosynthetic characteristics of the aquatic carnivorous plant *Aldrovanda vesiculosa*. *Aquat. Bot.* 59, 297–306.
- Adamec, L., 2000. Rootless aquatic plant *Aldrovanda vesiculosa*: physiological polarity, mineral nutrition, and importance of carnivory. *Biol. Plant.* 43, 113–119.
- Adamec, L., 2006. Respiration and photosynthesis of bladders and leaves of aquatic *Utricularia* species. *Plant Biol.* 8, 765–769.
- Adamec, L., 2007. Investment in carnivory in *Utricularia stygia* and *U. intermedia* with dimorphic shoots. *Preslia* 79, 127–139.
- Adamec, L., 2008a. Mineral nutrient relations in the aquatic carnivorous plant *Utricularia australis* and its investment in carnivory. *Fundam. Appl. Limnol.* 171, 175–183.
- Adamec, L., 2008b. The influence of prey capture on photosynthetic rate in two aquatic carnivorous plant species. *Aquat. Bot.* 89, 66–70.
- Adamec, L., 2009. Photosynthetic  $CO_2$  affinity of the aquatic carnivorous plant *Utricularia australis* (Lentibulariaceae) and its investment in carnivory. *Ecol. Res.* 24, 327–333.
- Adamec, L., 2010. Field growth analysis of *Utricularia stygia* and *U. intermedia* – two aquatic carnivorous plants with dimorphic shoots. *Phyton* 49, 241–251.
- Adamec, L., 2011a. Ecophysiological look at plant carnivory: why are plants carnivorous? In: Seckbach, J., Dubinski, Z. (Eds.), *All Flesh is Grass. Plant-Animal Interrelationships. Cellular Origin, Life in Extreme Habitats and Astrobiology*, Vol. 16. Springer Science+Business Media B.V., Dordrecht/Heidelberg/London/New York, pp. 455–489.
- Adamec, L., 2011b. Shoot branching of the aquatic carnivorous plant *Utricularia australis* as the key process of plant growth. *Phyton* 51, 133–148.
- Adamec, L., 2011c. Dark respiration and photosynthesis of dormant and sprouting turions of aquatic plants. *Fundam. Appl. Limnol.* 179, 151–158.
- Adamec, L., 2012. Why do aquatic carnivorous plants prefer growing in dystrophic waters? *Acta Biol. Sloven.* 55, 3–8.
- Adamec, L., Kovářová, M., 2006. Field growth characteristics of two aquatic carnivorous plants, *Aldrovanda vesiculosa* and *Utricularia australis*. *Folia Geobot.* 41, 395–406.
- Adamec, L., Pásek, K., 2009. Photosynthetic  $CO_2$  affinity of aquatic carnivorous plants growing under nearly-natural conditions and *in vitro*. *Carniv. Plant Newslett.* 38, 107–113.
- Borovec, J., Sirová, D., Adamec, L., 2012. Light as a factor affecting the concentration of simple organics in the traps of aquatic carnivorous *Utricularia* species. *Fundam. Appl. Limnol.* 181, 159–166.
- Draxler, G., 1973. Gaswechselformungen an *Utricularia vulgaris*. In: Ellenberg, H. (Ed.), *Ökosystemforschung*. Springer-Verlag, Berlin/Heidelberg/New York, pp. 103–107.
- Ellison, A.M., Adamec, L., 2011. Ecophysiological traits of terrestrial and aquatic carnivorous plants: are the costs and benefits the same? *Oikos* 120, 1721–1731.
- Friday, L.E., 1989. Rapid turnover of traps in *Utricularia vulgaris* L. *Oecologia* 80, 272–277.
- Helder, R.J., 1988. A quantitative approach to the inorganic carbon system in aqueous media used in biological research: dilute solutions isolated from the atmosphere. *Plant Cell Environ.* 11, 211–230.
- Jobson, R.W., Playford, J., Cameron, K.M., Albert, V.A., 2003. Molecular phylogenetics of *Lentibulariaceae* inferred from plastid *rps16* intron and *trnL-F* DNA sequences: implications for character evolution and biogeography. *Syst. Bot.* 28, 157–171.
- Kahara, S.N., Vermaat, J.E., 2003. The effect of alkalinity on photosynthesis-light curves and inorganic carbon extraction capacity of freshwater macrophytes. *Aquat. Bot.* 75, 217–227.
- Knight, S.E., 1992. Costs of carnivory in the common bladderwort, *Utricularia macrorhiza*. *Oecologia* 89, 348–355.
- Maberly, S.C., 1985. Photosynthesis by *Fontinalis antipyretica*. I. Interaction between photon irradiance, concentration of carbon dioxide and temperature. *New Phytol.* 100, 127–140.
- Maberly, S.C., Madsen, T.V., 2002. Freshwater angiosperm carbon concentrating mechanisms: processes and patterns. *Funct. Plant Biol.* 29, 393–405.
- Maberly, S.C., Spence, D.H.N., 1983. Photosynthetic inorganic carbon use by freshwater plants. *J. Ecol.* 71, 705–724.
- Madsen, T.V., Sand-Jensen, K., Beer, S., 1993. Comparison of photosynthetic performance and carboxylation capacity in a range of aquatic macrophytes of different growth forms. *Aquat. Bot.* 44, 373–384.
- Maier, R., 1973. Produktions- und Pigmentanalysen an *Utricularia vulgaris* L. In: Ellenberg, H. (Ed.), *Ökosystemforschung*. Springer-Verlag, Berlin/Heidelberg/New York, pp. 87–101.
- McDermott, M., Darnowski, D.W., 2002. Ecology of bladderworts in a unique site on the Eastern Shore of Maryland. *Carniv. Plant Newslett.* 31, 67–74.
- Moeller, R.E., 1978. Carbon-uptake by the submerged hydrophyte *Utricularia purpurea*. *Aquat. Bot.* 5, 209–216.
- Moeller, R.E., 1980. The temperature-determined growing season of a submerged hydrophyte: tissue chemistry and biomass turnover of *Utricularia purpurea*. *Freshwater Biol.* 10, 391–400.
- Nielsen, L.T., Borum, J., 2008. Why the free floating macrophyte *Stratiotes aloides* mainly grows in highly  $CO_2$ -supersaturated waters. *Aquat. Bot.* 89, 379–384.
- Pagano, A.M., Titus, J.E., 2004. Submersed macrophyte growth at low pH: contrasting responses of three species to dissolved inorganic carbon enrichment and sediment type. *Aquat. Bot.* 79, 65–74.
- Pagano, A.M., Titus, J.E., 2007. Submersed macrophyte growth at low pH: carbon source influences response to dissolved inorganic carbon enrichment. *Freshwater Biol.* 52, 2412–2420.
- Pechar, L., 1987. Use of acetone-methanol mixture for extraction and spectrophotometric determination of chlorophyll *a* in phytoplankton. *Arch. Hydrobiol.* 78 (Suppl.), 99–117.
- Pokorný, J., Ondok, J.P., 1991. Macrophyte Photosynthesis and Aquatic Environment. Academia, Prague.
- Richards, J.H., 2001. Bladder function in *Utricularia purpurea* (Lentibulariaceae): is carnivory important? *Am J. Bot.* 88, 170–176.
- Rutishauser, R., 1993. The developmental plasticity of *Utricularia aurea* (Lentibulariaceae) and its floats. *Aquat. Bot.* 45, 119–143.
- Sattler, R., Rutishauser, R., 1990. Structural and dynamic descriptions of the development of *Utricularia foliosa* and *U. australis*. *Can. J. Bot.* 68, 1989–2003.
- Sand-Jensen, K., Frost-Christensen, H., 1999. Plant growth and photosynthesis in the transition zone between land and stream. *Aquat. Bot.* 63, 23–35.
- Shipley, B., 2006. Net assimilation rate, specific leaf area and leaf mass ratio: which is most closely correlated with the relative growth rate? A meta-analysis. *Funct. Ecol.* 20, 565–574.
- Sirová, D., Borovec, J., Černá, B., Rejmánková, E., Adamec, L., Vrba, J., 2009. Microbial community development in the traps of aquatic *Utricularia* species. *Aquat. Bot.* 90, 129–136.
- Sirová, D., Borovec, J., Šantrůčková, H., Šantrůček, J., Vrba, J., Adamec, L., 2010. *Utricularia* carnivory revisited: plants supply photosynthetic carbon to traps. *J. Exp. Bot.* 61, 99–103.
- Sirová, D., Borovec, J., Píček, T., Adamec, L., Nedbalová, L., Vrba, J., 2011. Ecological implications of organic carbon dynamics in the traps of aquatic carnivorous *Utricularia* plants. *Funct. Plant Biol.* 38, 583–593.
- Taylor, P., 1989. The Genus *Utricularia*: a taxonomic monograph. In: *Kew Bulletin, Additional Series*, XIV.